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# Software package TotalXMR to support quantification of total content of X-nucleus from TD-NMR data

and an invitation to collaborate with us  
in order to solve many elemental analysis problems  
using your low- or high-field NMR equipment

## TotalXMR

Loads time-domain NMR data (FID, SE, QSE, CPMG, ...) from Bruker or other instruments and provides the user with a set of processing tools needed to

**assess the total content of the observed nuclides**

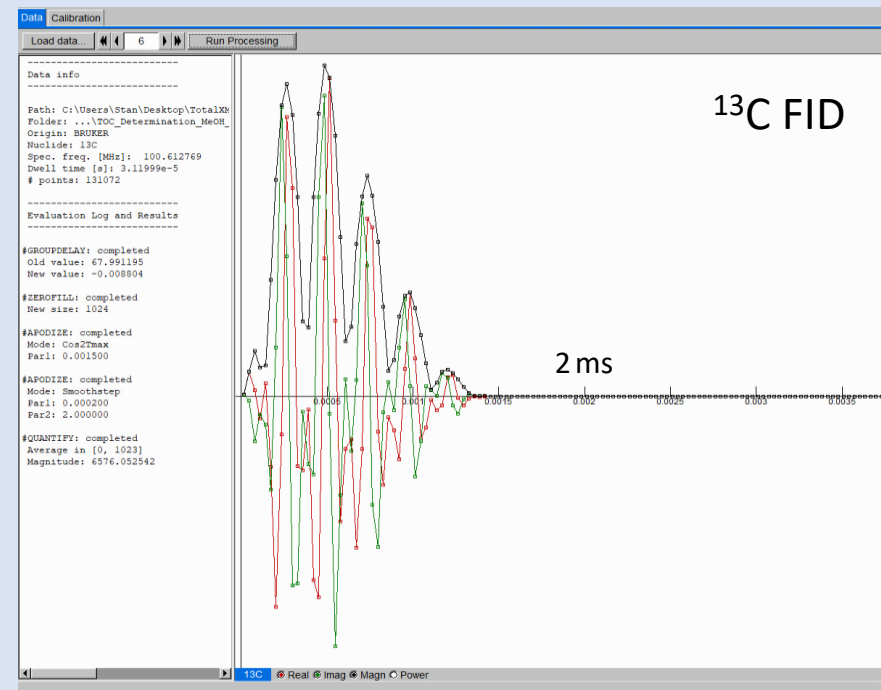
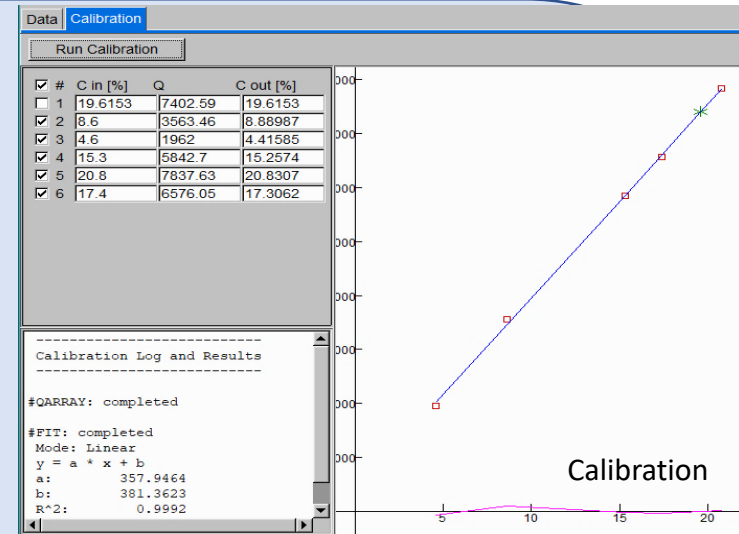
in an unknown sample, regardless of its composition.

Since NMR is intrinsically quantitative, such a task may appear trivial, but in practice it encounters a number of real-life obstacles that are listed in the next slide.

TotalXMR addresses many of these issues.

However, it needs to be always combined with a *data acquisition methodology* best suited for each NMR-active nuclide and each context.

*At present, there exists no universal data acquisition methodology for elemental analysis that would be applicable to every nuclide and every context.*



## What makes elemental analysis difficult?

Theoretically, the number of nuclei in the NMR coil is proportional to the FID signal intensity just after the excitation pulse. That sounds simple – so where is the problem? Some answers (though not all):

### Instrumental artefacts and limits:

- Group delays of digital receivers (see on the right)
- Limited sampling rates (=> SW limits)
- Receiver recovery time
- Probe ringing-down time

### Limits imposed by the nuclides (Mother Nature):

- Low sensitivity (S/N ratios) due to low gamma
- Low natural abundance

### Strong nuclear interactions (particularly in solids):

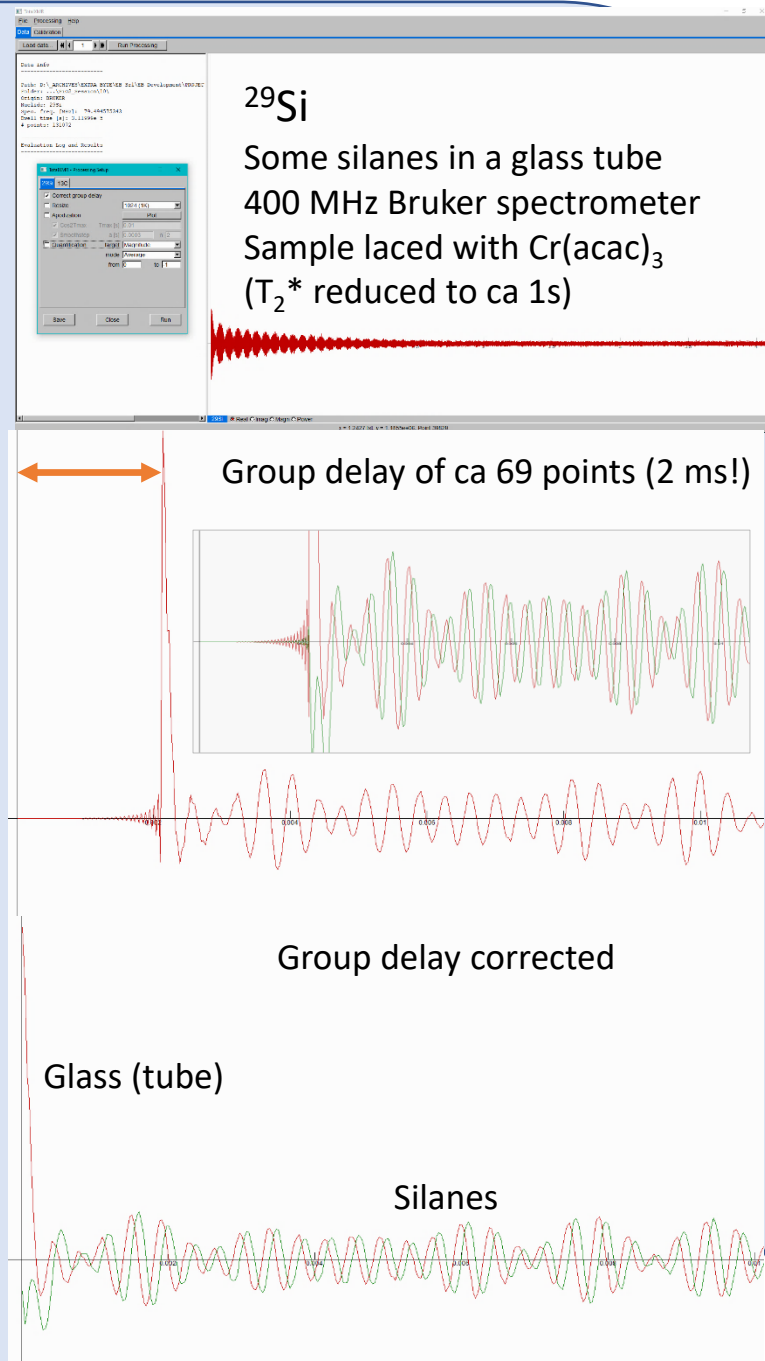
- Extreme quadrupole interactions (1 kHz – 5 MHz)
- Strong dipolar interactions (up to 120 kHz)

### Very long relaxation times (=> long acquisition times):

- Extremely long  $T_1$ 's (such as in many  $^{29}\text{Si}$  samples), particularly when combined with short  $T_2^*$  (solids).

### Sampling difficulties:

- Example: how to make a representative 1ml sample of, say, the soil in a certain agricultural area.



## Some salient features of TotalXMR

- Uses only a brief starting portion of TD-data.
- Automatically removes (known) artefacts.
- Can use both HR- or LR-NMR data.
- Is compatible with the use of relaxation agents to speed the assays.

### Special apodization methods:

- First suppress the very first (unreliable) points. Follow-up by isolating only a suitable starting portion of the data that makes the result independent of undesired details of chemical composition.

- If desired, however, leave in *some* chemical discrimination due to either  $T_2$  (solid/liquid) or to a specific shifts range (aliphatic/aromatic).

### Parameter settings:

- Very intuitive, user-friendly GUI.
- Allows separate settings for each nuclide.
- The setup can be saved and reused later.

### FID quantification selections

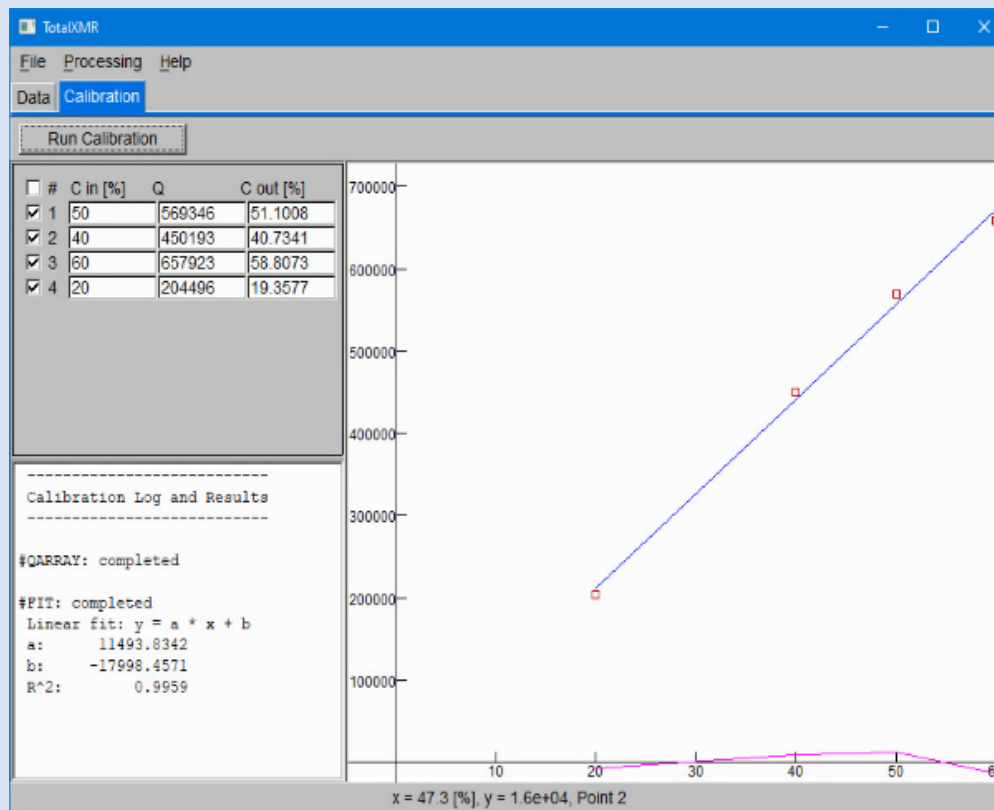
- Data can be windowed.
- Channel (magnitude/power/real).
- Various modes (average/maximum).

The image displays two windows from the TotalXMR software. The top window, titled 'TotalXMR', shows the 'Data' tab with a 'Calibration' sub-tab. It features a 'Data info' panel on the left with the following text: Path: C:\Users\manni\Desktop\Data; Folder: ...\SiO2\_Session\_10\40\; Origin: BRUKER; Nuclide: 29Si; Spec. freq. [MHz]: 79.494575349; Dwell time [s]: 3.11999e-5; # points: 191072. Below this is an 'Evaluation log and Results' section with entries for #GBCDELAY, #ZEROFILL, #APODIZE, and #APODIZE. To the right is a plot of the NMR data, showing a decaying oscillation. The bottom window, titled 'TotalXMR - Processing Setup', is for the 29Si nuclide. It has a 'Correct group delay' checkbox checked. Other options include 'Resize' (set to 1024 (1K)), 'Apodization' (set to Plot), 'Cos2Tmax' (checked, Tmax [s] = 0.01), 'Smoothstep' (checked, a [s] = 0.0003, n = 2), and 'Quantification' (checked, target = Magnitude, mode = Average, from = 0, to = -1). At the bottom are 'Save', 'Close', and 'Run' buttons.

## Calibration procedure:

- Load FID's of samples with either known or unknown concentrations.
- Set the desired parameters (or load them from a file)
- Input the concentrations or quantities of the calibration reference samples.
- Apply the Quantify command.

Automatically, all samples will be quantified using a **polynomial-regression**.



# Bring-home messages

Any NMR instrument that is capable of measuring an signal of a nuclide, can do so quantitatively, but it is not necessarily a trivial task.

The instrument may be low-field, high-field, table-top, full-size, field-cycling, any. TotalXMR needs to read the data that it produces, but we will see to that part.

Field homogeneity is not a problem, within very broad limits. Neither is the lock. Sensitivity, however, is an issue, some nuclei are viable only in high-field.

The acquired data needed for the evaluation are only the initial portions of time-domain signals (either FID's or more sophisticated ones). For good results, acquisition settings are important, and not necessarily matching your "standard" ones.

Many "tricks" regarding pulse sequences and/sample preparation are possible, we have some experience with that and we will be glad to collaborate with you.

Keep in mind that, once you buy a license, we become partners. The core of what we offer is not just the code: it is methodology + consulting + software-as-service.

**Nuclei you might be interested in:**  $^1\text{H}$ ,  $^2\text{H}$ ,  $^{13}\text{C}$ ,  $^{19}\text{F}$ ,  $^{31}\text{P}$ ,  $^{23}\text{Na}$ ,  $^{14}\text{N}$ ,  $^{15}\text{N}$ ,  $^7\text{Li}$ ,  $^{29}\text{Si}$ , ...

**Typical applications:**

**Si** in silicon glue, **C** in soil, **P** in meat, **N** in an animal feed, **Na** in cheese, **F** in a RedBull,

... ..

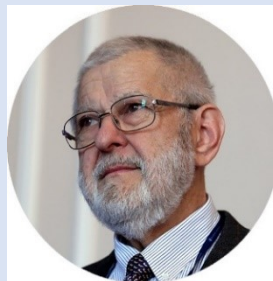
# Thank You for Your Attention!



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The team:



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In collaboration with

